

**S.G.A.P. ACACIA STUDY GROUP  
NEWSLETTER NO. 4  
MARCH 1969**

**Robert Coveny  
4 Churchill Crescent  
Concord NSW 2137**

We welcome three new members to the Group:-

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|-----------------------------------------------------------------------------------|---------------------------------------------------------------------|
| 1. <b>Mrs I B Armitage</b><br>"Timbarra"<br>410 Anembo Rd<br>Terry Hills NSW 2084 | 3. <b>Mr M Perry</b><br>27 Brazier St<br>Eaglehawk<br>Victoria 3556 |
| 2. <b>Mr K Rogers</b><br>Wulgulmerang<br>Victoria 3892                            |                                                                     |

Due to unavoidable commitments in November & December, it was not possible to produce the Newsletter at the regular time.

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As 1968 drew to a close, after twelve months leadership, it has been gratifying to see the Acacia Study Group continue to grow.

Several members have requested the names of all the people in the Group. These are given for the benefit of all members, especially those who have joined recently.

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|---------------------------------------------------------------------------------------|-------------------------------------------------------------------------------------------------|
| 1. <b>Mr Don Perrin</b><br>24 Pitt St,<br>Springwood<br>NSW 2777                      | 13. <b>Mr H L White</b><br>Nat. Library of Australia<br>Processing Branch<br>Canberra, ACT 2600 |
| 2. <b>Mrs E Chandler</b><br>20 Honiton Ave<br>Carlingford<br>NSW 2118                 | 14. <b>Mons. J Marnier-Lapostelle</b><br>91 Boulevarde<br>Hausman<br>Paris VIII E. France       |
| 3. <b>Mrs H Cumpston</b><br>24 Holmes Crescent<br>Campbell 2601<br>Canberra ACT       | 15. <b>Dr C Biccard Jeppe</b><br>PO Box 7240<br>Johannesburg<br>Republic of South Africa        |
| 4. <b>Miss M Pearce</b><br>"Dunolly"<br>Warne St<br>Katoomba NSW 2780                 | 16. <b>Mr Harry K Kurchian</b><br>287 Franklin Street<br>Reading 01867<br>Massachusetts, USA    |
| 5. <b>Mr W Payne</b><br>250 Picnic Pt Parade<br>Picnic Point NSW 2213                 | 17. <b>Mrs L Murphy</b><br>2 Kemsley Court<br>Hawthorn, Vic 3123                                |
| 6. <b>Mr N O'Donnell</b><br>Albion Crescent<br>Greensborough<br>Victoria 3088         | 18. <b>Mrs M J Monfries</b><br>5 Blacks Road<br>West Pennant Hills<br>NSW 2120                  |
| 7. <b>Mrs O McHaffie</b><br>"Carawatha"<br>Berringa Road<br>Park Orchards<br>Vic 3114 | 19. <b>Mrs J Hardy</b><br>132 Gloucester Rd<br>Hurstville,<br>NSW 2220                          |
| 8. <b>Mrs J B Stiller</b><br>"Hilldale"<br>Nagoorin, Qld 4680                         | 20. <b>Mr G W Althofer</b><br>Box 5<br>Dripstone NSW 2742                                       |

9.	<b>Mrs M Durbridge</b> Point Lookout North Stradbroke Island via Brisbane Qld 4184	21	<b>Mr K D Fairey</b> Box 1176 GPO Sydney NSW 2001
10.	<b>Mr &amp; Mrs J Simmons</b> 30 Ravenswood Rd Launceston Tasmania 7250	22	<b>Mr John Meredith</b> 7 Agar Steps Millers Point NSW 2000
11.	<b>Mr Clive Fisher</b> "Carinya" Old Bona Vista Road Warragul, Vic 3820	23	<b>Mr Des Nelson</b> PO Box 371 Alice Springs NT 5750
12.	<b>Miss P Males</b> 13 Gelding St Dulwich Hill NSW 2203	24	<b>Mr R Coveny</b> 4 Churchill Crescent Concord NSW 2137

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The following two books are recommended to members:-

**1. A Field Guide to Victorian Wattles** by F J C Rogers

An excellent book of 96 pages, with well defined line drawings, and a simple text which all members should be able to follow. In a recent letter from Mrs McHaffie, it is stated that this book is available from all the larger bookshops.

**2. Trees of New South Wales** by R H Anderson

A book of 510 pages, which should appeal to the country members. An excellent key is given in this book, for trees and shrubs over ten feet high. It is available from all the larger bookstalls, and would make a very good reference.

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This issue deals with insect pests which attack plants, both native and exotic. The control of insects is an important part of any gardening activity. Once the problem of insect infestation has been solved, a gardener can then expect to grow plants well if the right conditions are available for that particular plant.

In this issue, the main insect pests that attack cultivated plants in general are dealt with – giving the symptoms that the various insects cause upon plant growth, and also chemicals that could be used for their control.

In a future issue it is intended to write about diseases which attack cultivated plants, both native and exotic, also to mention the storage of seed, giving treatment which could be employed to ensure that seed will be safe from infection from insect pests, especially weevils.

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## INSECT PESTS

**Plague Thrips** – “A native to Australia, which may occur in plague numbers in spring and early summer, when climatic conditions are favourable to its development. This species is present every year in limited numbers, but serious spring outbreaks have been found to follow an autumn or winter in which the rainfall has been above average.

The adult thrip feeds in blossoms of weeds, fruit trees, vegetables and ornamental garden plants. Injured flowers may turn brown and fall prematurely. The thrips enter the opening blossoms and feed on the petals.”

### Control

D.D.T. emulsion 0.1% or  
Malathion emulsion 0.05% or

White Oil 10 fluid oz, Nicotine Sulphate 1 fluid oz, + water 4 gallons, or  
2% D.D.T. dust, or  
2% B.H.C. dust, or 5% dust.

When using D.D.T. emulsion, spray early in the morning, late in the afternoon, or even at night, when bees are not active.

**White Wax Scale** – “A pest of citrus orchards, also infests many native and introduced plants, such as Blackthorns, Lillypilly, Gardenia, Pittosporum, Persimmon, Guava, Eriostemon etc.

On removing the wax covering, the soft reddish insect is found below. The larvae or “crawlers”, emerge from the eggs beneath the parent scale, mainly from November to January, and settle along the midribs and veins of the leaves. About 5 or 6 weeks later, they crawl back to the twigs, settle permanently in position and secrete the wax cover, increase in size, and about 10 months later, lay eggs.”

### **Control**

White Oil 1:40

White Oil 1:80 + Malathion 0.1% dispersible powder or

White Oil 1:80 + Triton 0.05% d.p. or

White Oil 1:80 + Ethion 0.05% d.p. or

White Oil 1: 80 + Gusation 0.05% d.p.

Spray mid-Dec to early January when larvae on leaves after most have hatched, and before wax production commences on stems. Thorough spraying on leaf surfaces is essential.

Washing soda 10lbs per gallon or

Soda ash 3½ lbs per gallon.

Before mid-Feb to kill “Peak-Stage”.

**Plant Bugs:** “Among some of the native species of plant bugs that at times become sufficiently numerous to cause damage to cultivated plants is the Crusader Bug. These bugs are present in small numbers in most seasons. Under natural conditions, they live on native trees and shrubs and on various weeds, but in seasons favourable for their development they reach plague numbers, and it is at such times that cultivated plants are infected.

All bugs live by piercing the plant tissue and sucking up the sap, and increase in size by a series of moults, until the adult winged stage is reached. They feed on Wattles, Eucalypts, Cassias etc.”

### **Control**

D.D.T. 0.1% spray, or

Malathion 0.075% spray, or

Lindane 0.075% spray, or      Spray when observed

Rogor 0.04% spray, or

Malathion 5% dust.

**Leafhoppers:** “These bugs range from very small to medium sized insects, and are often very abundant. They pass the whole of their life-cycle above ground, on stems, leaves, or grasses, and feed entirely on plant sap. Many of the smaller species occur in large swarms, and cause considerable damage to trees, shrubs, and other cultivated plants. A typical example is the Passion Vine Leaf-hopper, which feeds naturally on various native trees and shrubs. They are capable of both hopping and flying when disturbed.”

### **Control**

Nicotine sulphate 1 fluid oz. in 4 gallons water, or

D.D.T. 0.1% spray, or

Malathion 0.05% spray, or

Rogor 0.04% spray, or

Lindane 0.03% spray.

Spray when immature stages present. Spray must also be applied over soil surface, after treating infected plant.

**Aphids:** “Cultivated ornamentals serve as primary or secondary food plants of many species of aphids, and because of their small size and often concealed positions on plants, they are frequently overlooked, until a heavy infestation has developed. They are all small soft-bodied insects, which may be found on the under-sides of leaves, around flower buds, and on the young shoots and bark, or on the roots of various plants. Others again may live in galls. Some species, by means of special glands, secrete varying amounts of a white waxy substance, which may be either powdery or woolly, and this may cover their bodies. Most species, however, are without any protective covering. They are of various colours, depending on the species, and sometimes on the food plants. Aphids possess sucking beaks, and feed by puncturing the plant tissues and extracting the sap. When numerous, they may dry up and curl leaves, distort the buds and flowers, and weaken the entire plant.

Some aphids cause scars and gall-like swellings to form on the limbs, twigs and roots of the plants. Many are capable of transmitting plant virus diseases.

Most species excrete quantities of “honey dew”, a sugary substance, which adheres to, and disfigures the foliage on which it falls. A black or “sooty” mould, which develops on the “honey dew”, adds to the disfigurement. Ants and flies are attracted to this secretion and feed upon it. The honey dew of some aphids may cause a “burning” of the foliage upon which it falls.

Some aphids, during the autumn and early winter, lay eggs which overwinter. These eggs hatch in the spring, and a generation of wingless aphids develops. This first spring generation gives birth to living young, and they in their turn produce further living young, so that generation after generation, all the individuals of which are either winged or wingless females, continue to develop in this manner until the approach of autumn, when wingless egg laying females and winged females (or wingless in some species) make their appearance. The females of this generation, after mating, lay the overwintering eggs.

Aphids are attacked by a large variety of natural enemies, including parasitic wasps and predacious ladybirds and their larvae, and syrphid fly and lacewing fly larvae. These natural enemies often control aphid infestations naturally, but if the natural enemies are killed out by injudicious use of sprays, the aphid infestation may increase considerably a short time after spraying. Ants attending aphids (to feed on honey dew) tend to repel the small wasp parasites and control of ants is often followed by a marked increase in natural control of parasites.”

## Control

1. Nicotine sulphate 1 fluid oz, soap 2 oz, water 4 gallons.
2. 2½% Nicotine Dust.
3. Malathion spray 0.05%
4. Metasisosystex spray 0.02%
5. Ekatin 0.025% spray
6. Sayfos 0.01% spray

Spray when observed.

Root feeding aphids – Malathion, Lindane or Nicotine sulphate.

**Scale Insects:** “ Many ornamentals are attacked by a number of species of scale insects. These various species belong to a number of families in the super-family Coccoidea, which also includes the mealy bugs, some gall formers. All of these forms are characterised by the fact that the female is always wingless, often stationary and degenerate, while the male, if present, is winged, with one pair of wings only.

In some species, the males are rare or absent, and reproduction may be entirely parthenogenetic. One species, Cottony Cushion Scale, is a functional hermaphrodite.

The scale insects are the most abundant and familiar forms, and many of them are of considerable economic importance. Many have fairly wide host ranges, although some are restricted to a single host plant or a group of closely related plants. The adult female is degenerate, stationary, and protected by a scaly covering (Armoured Scales, Family Diaspididae) or by a thickened integument or thick coating of wax (Family Coccoidea, the Soft and Wax scales).

Most scale insects secrete honeydew, which serves as food for ants, and also acts as a medium for the growth of sooty mould fungi.

A typical example of the armoured scale is the Citrus Red Scale. Heavy infestations of this scale have been seen on various ornamentals, including Acacia, Euonymus, Ligustrum, Rose and Salix.”

The following are the most common armoured scales which attack cultivated natives and exotics:-

1. Yellow Scale. The female scale is flat, circular, translucent, the yellow adult body showing through it. The male scale is yellow, grey and elongate. Many hosts.
2. Oleander Scale. The female scale is white or pale grey, and circular. The male scale is similar in colour and slightly oval. Hosts include Cymbidiums and Acacias.
3. Greedy Scale. Adult female scale grey, markedly convex, with a strongly lipped-over appearance. Male scale unknown. Wide host ranges includes Acacia species.
4. Rossi's or Flat Backed Scale. Female scale is quite flat, circular, very dark brown or black. Male scale slightly elongate, oval, dark brown. Hosts include Banksia, Palms, Abutilon, etc.
5. Mussel Scale. Adult female scale elongate, slender, variable and grey to purplish in colour. This scale is apparently parthenogenetic. Wide host range.
6. White Palm Scale. Female scale is 1/10 of an inch long, white, elongate and pear-shaped in outline. The male scale is much smaller, elongate, narrow and usually covered with white cottony secretion. Hosts include Palms, Ceratopetalum, Melaleuca, Leptospermum, Waratah.
7. Aspidistra Scale. Adult female scale elongate, quite thick, definitely brown, male scale elongate, parallel sides, white felted and bearing three longitudinal ridges. Hosts include Cassia, Cymbidiums, Ferns.

The following are the most common soft scales which attack cultivated natives and exotics:-

1. Citrus White Wax Scale. Hosts include Lillypilly, Pittosporum, Native Blackthorn etc.
2. Indian White Wax Scale. This introduced species is of minor economic importance, occurring on a number of garden and native plants.
3. Pink Wax Scale. Smaller than the white wax scale, its colour is rose-red or pale greyish pink, and it is about 1/8 of an inch long. The wax is harder and more crystalline. Hosts include Lillypilly, Pittosporum, Leptospermum, Tree Fern.
4. Nigra Scale. Adult female scale is oval to slightly elongate, very convex, smooth and shining, reddish-brown to black in colour, and about 4mm long. Hosts include Pittosporum.

Various other families in the super family Coccoidea are of economic importance. The family Margarodidae includes some of the largest native species of mealy bugs, in the genera Monophloioies and Callipappus found on Melaleuca and Eucalyptus, the ground pests Margaredes, which are subterranean, and feed on plant roots, and the Cottony Cushion Scale, Icerya purchasi. This native species is so called on account of the white fluted mass of woolly secretion in the egg sac, which is formed by the reddish-brown female. In NSW it infests a variety of hosts, including wattle, citrus, climbing rose and laburnum. It can be controlled by the ladybird *Rodolia cardinalis*, one of the two native parasites which largely keep the pest in check in this state.

## Control

Armoured and Soft Scales

White Oil 1:40  
White Oil 1:80  
White Oil 6 fluid oz, Malathion 25% w.p. + water 3 gallons.

Spray during peak emergence of crawlers, often indicated by ant activity.

Wax Scale

White Oil 1:40, or  
White Oil 6 fluid oz, Malathion 25% w.p. 20oz, water 3 gallons or  
Washing Soda 1½ lb or 9oz soda ash to 4 gallons of water , plus  
white oil 1:40 or  
Soap at 4oz in 2 gallons water.

Spray when young crawlers on leaves (mid to end of December for white wax), December – February for pink.

Cottony Cushion Scale

White Oil 1:40, or  
White Oil 6 fluid oz, Malathion 25% w.p. 20oz, water 3 gallons.

Spray when necessary, second application 4-6 weeks later may be necessary.

**Mealy Bugs:** These insects, which are related to the scale insects, are covered with a white mealy wax, from which they have derived their popular name. Many of the species, in addition to their mealy covering, also have a characteristic fringe of white filaments around the margins of their bodies.

Native mealy bugs do not appear to be of great economic importance, but members of the introduced species are amongst the most serious pests of plants. Heavy infestations may occur on various cultivated trees, shrubs and ornamental plants, including Daphne, Gardenia, Orchids and Ferns. Leaves and roots of various grasses and clovers may be attacked.

Mealy Bugs are most numerous where the climate is moderate to warm with moist conditions. The temperature and humidity usual in glasshouses and ferneries, or where plants are grown indoors, are very favourable for their development, and under these conditions they increase in numbers very quickly.

The sheaths about the bases of leaves and the undersides of leaves, the lower parts of fern fronds, and the young growth of flower buds of various plants, are favoured feeding places. Infestations in these parts often pass unnoticed until the mealy bugs have increased to enormous numbers, or the plants begin to wilt.

Mealy Bugs feed by piercing the plant tissues and sucking up the sap. They secrete quantities of "honeydew", which falls upon the foliage and other parts of the plants, and in this sugary secretion sooty moulds develop. Ants are attracted by the "honeydew" and feed upon it, and to some extent prevent the mealy-bugs being attacked by natural insect enemies.

Where ants are present, they may assist in the spreading of root-feeding mealy bugs, as they construct galleries, along or among the roots of the plants, providing the mealy-bugs with ready access to fresh roots.

## Control

1. Nicotine sulphate 1 oz, white oil 8 oz, water 4 gallons.
2. White Oil 6 fluid oz + Malathion 25% w.p. 20 oz, plus water 3 gallons.
3. Malathion 5% dust.
4. Metasystox 1:500.
5. Methylated Spirits.

Spray when observed. Ant control, using 2% Chlordane, is often worthwhile.

These notes, dealing with insects and their control, were taken from Tech. notes, and will be completed in the next Newsletter.

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It was my intention to produce an article dealing with the Acacias seen on my Central Australian tour, but unfortunately have not had time to do so.

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Three articles of interest from **Mr John Meredith**, are reproduced below.

## WATTLE DAY

"In my youth, Wattle Day was an outstanding annual event. Every pupil turned up at school wearing a sprig of our national emblem (generally *A. baileyana* or *A. decurrens*), assembly was given over to a talk about wattle, and we sang songs, such as "The Graceful Swaying Wattle". In recent years, the festival has been sadly neglected, especially in the cities and larger towns. It was encouraging then, to hear that the Gould League of Bird Lovers has merged with the Junior Tree Gardens, to form a new organization known as the Gould League, and that they are going to devote some of their activities toward re-popularising Wattle Day.

Wattle Day originated in an idea put forward to J H Maiden at a Sydney public meeting on the 30<sup>th</sup> August, 1909, and August 1<sup>st</sup> 1910 was fixed upon as the date for the first Wattle Day. Since then the festival has been observed all over Australia on 1<sup>st</sup> of August or 1<sup>st</sup> of September, according to the flowering period in each State. At the time of the public meeting, Maiden was curator of the Sydney Botanic Gardens. Also present at the meeting were Mrs J Kettlewell and Mrs Clunies-Ross.

The original idea was to celebrate the embodiment of the Australian national sentiment in a native flower, and the setting apart of a day for its celebration.

I feel that our Group could and should help in re-establishing Wattle Day as a popular National Day, and suggest several ways in which we could do this:-

1. Encourage the cultivation of the Acacia as a garden plant.
2. Popularise the use of the Acacia as a cut flower, and to this end, discover and make known methods of preserving the freshness of the cut acacia blooms in water eg burnt or scalded stems ?, additives such as sugar, honey or aspirin to the water?
3. Campaign for the August Bank Holiday to be known as Wattle Day, and to be gazetted as a general public holiday.
4. Campaign for the establishment of a "Wattle Festival" to be held on the holiday, and which, as befitting our national emblem, could be held in Canberra."

John Meredith

## GERMINATION OF ACACIA SEED

"This summer I planted more acacia seed than I have ever tried to raise before. The results were, in the main, encouraging. Most species gave more than 50% germination, a few yielded only one or two seedlings, and some, as usual, failed to germinate altogether. Using approximate figures of 50% for good, 30% for fair, and nil for poor, the following list will give some idea of my successes and failures:-

Good:- *A. podalyriifolia*, *A. spectabilis*, *A. pycnantha*, *A. impressa*, *A. jibberdingensis*, *A. pyrifolia*, *A. tetragonophylla*, *A. glaucescens*, *A. rubida*, *A. vestita*, *A. decora*, *A. fennifolia*, *A. victoriae*, *A. tanumbirinensis*.

Fair: *A. giraffii*, *A. cowleana*, *A. georginae*, *A. undulifolia*, *A. doratoxylon*.

Poor: *A. acuminata*, *A. drummondii*, *A. dunnii*, *A. xiphophylla*, *A. alata*, *A. mitchellii*, *A. merrallii*, *A. murrayana*, *A. iteaphylla*, *A. karoo*, *A. pubescens*, *A. albida*, *A. aspera*, *A. baeuerlenii*, *A. brunioides*, *A. galpinii*, *A. pugioniformis*, *A. subulata*, *A. estrophiolata*.

These figures apply only to germination, and not to success in raising the plants. Owing to the unusually dry and windy conditions I lost a lot of seedlings due to dehydration and damping off, as I am only able to water them at weekends."

John Meredith

## ACACIAS FAILING TO SET SEED

"At Balmoral in the Southern Highlands, there was a profuse blooming of acacia this year, and I looked forward to a good harvest of seed. When the time arrived to collect seed, however, I found that many species had failed to set seed, and some had only a sparse scattering of seed pods.

There are two possible causes for this phenomenon. Firstly, the prevailing south-westerly winds, this year dry and very strong and over a prolonged period – from April to December. I feel that this wind could have caused dehydration and wilting of the flowers before the bees were able to pollinate them. The other possible cause is the drought, this is the fourth year in which the annual rainfall was only about half the usual average.

Species which had only a sparse setting of seed are: *A. myrtifolia*, *A. fimbriata*, *A. decurrens*, *A. baileyana* and *A. baileyana* var. *viridifolia*, *A. linifolia*, *A. decora*, *A. ulicifolia*.

Those which appeared to yield the usual number of seed pods were: *A. pycnantha*, *A. elata*, *A. suaveolens*."

John Meredith

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The first part of my check list on Australian Acacias appears in this issue. The South Australian list is the only one which has been checked by a botanist and returned to me. No synonyms have been given, as there still appears to be some confusion in the correct application of names.

## SOUTH AUSTRALIAN ACACIAS

1. *A. acanthoclada* F. Muell.
2. *A. acinacea* Lindl.
3. *A. anceps* DC, var. *anceps*  
*A. anceps* DC, var. *angustifolia* Benth.
4. *A. aneura* F. Muell. ex Benth. var. *aneura*  
*A. aneura* F. Muell. ex Benth. var. *latifolia* Black
5. *A. argyrophylla* Hook.
6. *A. armata* R. Br. ex Ait.
7. *A. barattensis* Black
8. *A. basedowii* Maiden
9. *A. beckleri* Tindale
10. *A. brachybotrya* Benth.
11. *A. brachystachya* Benth.
12. *A. burkittii* F. Muell. ex Benth.
13. *A. bynoeana* Benth. var. *latifolia* Black (not valid)
14. *A. calamifolia* Sweet ex Lindl.
15. *A. calcicola* Forde & Ising
16. *A. cabbagei* R. T. Baker
17. *A. cana* Maiden \*
18. *A. carnei* Maiden
19. *A. colletioides* A. Cunn. ex Benth.
20. *A. continua* Benth.
21. *A. coriacea* DC. \*
22. *A. coronalis* Black \*
23. *A. cyclophylla* Schldl.
24. *A. cyclops* A. Cunn. ex G. Don
25. *A. cyperophylla* F. Muell. ex Benth.
26. *A. dictyophleba* F. Muell.
27. *A. dodonaeifolia* Willd. ex Spreng.
28. *A. enterocarpa* R. V. Smith
29. *A. erinacea* Benth.
30. *A. estrophiolata* F. Muell.
31. *A. euthycarpa* (Black) Black
32. *A. farinosa* Lindl.
33. *A. farnesiana* (L.) Willd.
34. *A. gilesiana* F. Muell.
35. *A. gillii* Maiden et Blakely
36. *A. gracilifolia* Maiden et Blakely
37. *A. hakeoides* A. Cunn. ex Benth.
38. *A. havilandii* Maiden
39. *A. imbricata* F. Muell.
40. *A. iteaphylla* F. Muell. ex Benth.
41. *A. kempeana* F. Muell.
42. *A. ligulata* A. Cunn. ex Benth.
43. *A. lineata* A. Cunn. ex G. Don
44. *A. linophylla* W. V. Fitzg.
45. *A. loderi* Maiden
46. *A. longifolia* (Andr.) Willd. var. *longifolia*  
*A. longifolia* (Andr.) Willd. var. *sophorae* (Labill.) F. Muell.
47. *A. maitlandii* F. Muell.
48. *A. mearnsii* De Wild.
49. *A. melanoxyton* R. Br.
50. *A. menzellii* Black
51. *A. merrallii* F. Muell.
52. *A. microcarpa* F. Muell. var. *linearis* Black  
*A. microcarpa* F. Muell. var. *microcarpa*
53. *A. mitchellii* Benth.

54. *A. montana* Benth.
55. *A. murrayana* F. Muell. ex Benth.
56. *A. myrtifolia* (Sm.) Willd. var. *angustifolia* Benth.  
*A. myrtifolia* (Sm.) Willd. var. *myrtifolia*
57. *A. notabilis* F. Muell.
58. *A. oswaldii* F. Muell.
59. *A. oxycedrus* Sieb. ex DC.
60. *A. papyrocarpa* Benth.
61. *A. peuce* F. Muell. \*
62. *A. pinguifolia* Black
63. *A. pravifolia* F. Muell.
64. *A. prolifera* Black
65. *A. pycnantha* Benth.
66. *A. quornensis* Black
67. *A. ramulosa* W. V. Fitzg.
68. *A. randelliana* W. V. Fitzg.
69. *A. retinodes* Schlecht. var. *oraria* Black et Bardley  
*A. retinodes* Schlecht. var. *retinodes*  
*A. retinodes* Schlecht. var. *uncifolia* Black
70. *A. rhetinocarpa* Black
71. *A. rhigiophylla* F. Muell. ex Benth.
72. *A. rigens* A. Cunn, ex G. Don.
73. *A. rivalis* Black
74. *A. rivularis* Black
75. *A. rotundifolia* Hook.
76. *A. rupicola* F. Muell. ex Benth,
77. *A. salicina* Lindl.
78. *A. sclerophylla* Lindl. var. *lissophylla* Black  
*A. sclerophylla* Lindl. var. *sclerophylla* Lindl.
79. *A. sessiliceps* F. Muell.
80. *A. signata* F. Muell.
81. *A. sowdenii* Maiden
82. *A. spilleriana* J. E. Brown
83. *A. spinescens* Benth.
84. *A. stenophylla* A. Cunn. ex Benth.
85. *A. stricta* (Andr.) Willd.
86. *A. strongylophylla* F. Muell.
87. *A. suaveolens* (Sm.) Willd.
88. *A. tarculensis* Black
89. *A. tenuior* Maiden
90. *A. tetragonophylla* F. Muell.
91. *A. trineura* F. Muell.
92. *A. validinervia* Maiden et Betche
93. *A. verniciflua* A. Cunn.
94. *A. verticillata* (L'Her) Willd. var. *latifolia* Benth. \*
95. *A. victoriae* Benth.
96. *A. vomeriformis* A. Cunn. ex Benth.
97. *A. wattsiana* F. Muell. ex Benth.
98. *A. wilhelmiana* F. Muell.